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Methods of breaking seed physical dormancy and germination in native species of Alhagi graecorum **Boiss (Al-Agool)**

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ABSTRACT **Keywords:** Alhagi graecorum is a plant belonging to the Fabaceae family and grows as wild perennial shrubby Fabaceae family Alhagi graecorum species. Seeds of this species possess seed physical dormancy and need to be removed to enhance Physical dormancy germination, breaking of dormancy treatments were imposed on seeds to improve germination. Breaking dormancy Treatments include scarification with sulfuric acid (H₂SO₄) for 10, 20, 30 and 40 minutes. The results Scarification showed highly significant difference between control and all the treatments of germination percentage Germination (GP) was 96%, 96%, 97% and 97%, respectively, while recorded in control only 12%. As for the mean daily germination (MDG), the results revealed that, there was a significant differences between all treated seeds and control, which was the fastest and most effective seeds germination on the third day of sowing were 84, 89, 90, 92 % respectively, while in control was 0 %. Mean germination time (MGT) decreased in all treated seed but statistically same while significantly different from the control. The minimum time was recorded for 20 and 30 minutes was 3.14 and 3.14 days respectively.

We conclude from the results obtained that, the use of concentrated sulfuric acid 98% achieved the highest GP and MDG and the lowest MGT. This is the efficient method of breaking seeds dormancy and germination for native species of Alhagi graecorum.

طرق كسر السكون الفيزيائي والإنبات لبذور نبات العقول المحلى Alhagi graecorum Boiss

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الكلمات المفتاحية: الملخص العائلة البقولية نبات العقول (Alhagi graecorum) ينتجي إلى العائلة البقولية، شجيرى عشبي معمر. تمتلك بذور هذا النوع سكونًا فيزيائيا، ولتعزيز الإنبات بحاجة إلى إزالته، ولكسر السكون لتحسين الإنبات، اشتملت معاملات العقول السكون الفيزيائي الخدش باستخدام حمض الكبريتيك المركز لمدة 10، 20 ، 30 و 40 دقيقة. أظهرت النتائج وجود فروق كسر السكون معنوبة عالية في كل المعاملات في نسبة الإنبات و التي بلغت 96٪ ، 96٪ ، 97٪ و 97٪ على التوالي، بينما سجلت الخدش في الشاهد 12٪ فقط. أما متوسط الإنبات اليومي، فقد أوضحت النتائج وجود فروق ذات دلالة إحصائية بين الإنبات جميع البذور المعاملة بالحمض مع الشاهد، حيث كان إنبات البذور فيها الأسرع والأكثر فاعلية في اليوم الثالث من البذر فكانت 84 ، 89 ، 90 و 92/ على التوالى ، بينما في الشاهد 0/. أما بالنسبة لمتوسط زمن الإنبات، انخفض في جميع البذور المعاملة بالحمض، بينما لم تظهر وجود اختلافات معنوبة فيما بينها، وكان الاختلاف معنوباً مع الشاهد. تم تسجيل الحد الأدني لزمن الإنبات عند المعاملة بحمض الكبريتيك لمدة 20 و 30 دقيقة فقد كانت 3.14 ، 3.14 يومًا على التوالي. نستنتج من النتائج التي تحصلنا عليها، أن استخدام حامض الكبريتيك المركز % 98٪ أعطى أعلى نسبة إنبات و أعلى متوسط يومي للإنبات و أدنى متوسط لزمن الإنبات. لذلك تعتبر هي الطريقة الفعالة لكسر سكون البذور

و إنبات هذا النوع المحلي لبذور نبات العقول (Alhagi graecorum).

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Introduction

Fabaceae (Leguminosae) is a family with high value of use for food, medicine, forage, ornamental and restoration ecology purposes. One obstacle to the use and management of many legume species is the presence of physical seed dormancy [1] The genus Alhagi belongs to the family Fabaceae (Leguminosae) and is consisted of about nine species ^[2] The Latin name is derived from the Arabic name Alhag, meaning the old wise man ^[3]. Alhagi species are ordinarily used in folk medicine as remedies to treat rheumatism, bronchitis, ulcers, liver disorders and jaundice, urinary tract diseases, asthma and gallbladder problems, recent in vivo and in vitro biological activity studies on Alhagi species revealed their antibacterial, antifungal, antioxidant, antiproliferative, hepatoprotective spasmolytic, and ureter-relaxing effects^[4]. Alhagi graecorum Boiss, (Camel thorn), known locally as, Al-Agool, is an indigenous plant, shrubby evergreen perennial herb, branched, with rigid spiny twigs, long roots and wide ecological amplitude that allows it to withstand harsh environments, drought tolerant. This species is widely distributed in Murzuq, Ghat, Barkat, Wadi Al Ajal and Al Jaghbub, and grows naturally in xeric successfully and intensively in dry, saline soil, making its in ecosystem desert areas in southwestern Libya, and produces a satisfactory vegetation cover that protects soil from erosion, and it can be highly useful for prevention of land degradation (Fig. 1). It is currently used as a forage for Camels, goats and sheep in a southwest desert regions of Libya (Fig. 2).



Fig. 1: Alhagi graecorum Boiss (Habit)



Fig. 2:Alhagi graecorum, harvested, collected and sold as livestock forage

Dormancy is a term used to describe a seed that fails to germinate under favorable conditions at a specified time ^[5], it's can be biologically described as: "The absence of germination of an intact, viable seed under germination favoring conditions with a specific time lapse ^[6]. Dormancy acts in mimetic-seeded species as an exaptation to reduce seed deterioration. Dormancy is of great importance in an evolutionary perspective and as well in fitness [7],[8], and also important in aspects of dispersal and as a mechanism for delaying seed germination until it has been spread to new areas ^[9]. Seed dormancy is defined as the failure to germinate under such favorable conditions, although the viable seed [10], [11], and is a temporary failure or block of a viable seed to complete germination under physical conditions that normally favor this ^{[12],[13]}, it's a block to the completion of germination of an intact viable seed under favorable conditions ^[14], also it is the resting period of seed after physiological maturity and also an adoption mechanism to overcome stress conditions ^[15], as well it's important component of plant fitness that causes a delay germination until the arrival of a favorable growth season [16], It's an innate seed property that defines the environmental conditions in which the seed is able to germinate [14],[17]. Seeds of plant species with physical dormancy are known in 17 families of angiosperms ^[18], and several types of specialized structures ('water gaps') have been found in 12 of the 17 families. Physical dormancy is present in species of at least 15 angiosperm families, including Fabaceae, Malvaceae, Convolvulaceae, Chenopodiaceae, Cannaceae, and Liliciae. In some of these species, seed coat impermeability may delay germination for several years. Physical dormancy is caused mainly by impermeable seed coats that prevent water uptake. The Majority of Leguminosae species have hard and water impermeable seed coats, that inhibits seed germination and causes dormancy. Although hard of the seed coat is a structure which protects the embryo from mechanical effects, it has a negative impact on germination^[19]. The family of Fabaceae (Leguminosae) has a large number of species with physically dormant seeds [20]. Within the Fabaceae family, the structure associated with the breaking of dormancy is usually the lens^[21]. The objective of this experiment was to determine the effective of sulfuric acid (conc.H2SO4) on breaking dormancy and promoting germination of Alhagi graecorum seeds

Materials and methods

Mature pods of *Alhagi graecorum* were collected from A shrubs growing in different places of the Murzuq region - Libya (N: 55° 25", E: 55° 13", 449 m). (Fig. 3). Seeds were removed from the pods immediately (Fig. 4,5), and stored in glass bottles at room temperature



Fig. 3: Map of Libya showing the location of Murzuq



Fig. 4: Mature pods of Alhagi graecorum



Fig. 5: Seeds of Alhagi graecorum

Chemical scarification

100 Seeds were counted per treatment and soaked separately in sulfuric acid (conc. H_2SO_4 98%) for various time intervals 10, 20, 30, and 40 minutes (min.), to evaluate the time required to break the dormancy of the seeds, scarified seeds were rinsed several times in clean distilled water after the treatment with acids to remove any trace of acid, after rinsing, seeds were allowed to dry on blotter paper at the laboratory temperature, before being placed in Petri dishes. Untreated seeds were used as control. Germination tests, were undertaken with 5

replicates for H₂SO₄, each replicate contained 20 seeds placed in 9 cm sterilized Petri dishes lined with double layered of Whatman No. 1 filter paper. The papers were moistened with 5 mL distilled water and covered-up with their respective covers, and added distilled water was necessary, to prevent seeds from drying out. Afterward, the dishes were incubated in dark at 25 °C. Seed germination was counted and recorded daily until no further germination occurred. The criterion for germination was visible radicle protrusion from the seed coat ^[22]. At the end of the incubation time, the following parameters were assessed. Germination percentages were calculated using the following equation.

Germination percentages (GP) = $\frac{G}{N} \times 100$

Where G = Total number of seeds that germinated.

N = Total number of seeds in the Petri dish. ^[23]. Mean daily germination (MDG), an index of daily germination rate, was determined on the basis of the following equation ^{[24],[25]}.

MDG =
$$\frac{FGF}{D}$$

FGP is the final germination percent. And D is experiment period. Mean Germination Time (MGT day)

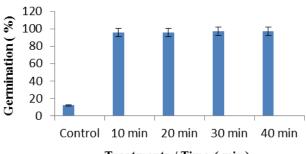
MGT = $\Sigma f \cdot x / \Sigma f$ f = Seeds germinated on day x ^[26].

Statistical analyses.

Statistical analysis of data and the treatments means were tested by the one way analysis of variance (ANOVA). Mean comparison was performed using Minitab least-significant difference (LSD) method P < 0.05 considered significant Values were expressed as means \pm SD (standard deviation) for five replicates in each of the independent experiments.

Results and Discussion

From an ecological perspective, dormancy is an important survival mechanism that favors the propagation and dissemination of seeds to establish plant populations. Because specific conditions are required to break dormancy, it may favor germination and seedling emergence under more favorable conditions. In generally treated seeds recorded positive response on germination, the results revealed treatments including scarification with sulfuric acid (H_2SO_4) for different soaking durations were significant effects in all aspects of germination tests. From the results obtained in (Fig. 6) seeds scarified in H_2SO_4 at different durations for 10, 20, 30 and 40 minutes produced high germination percentages (GP) 96%, 96%, 97%, 97% that were highly significant differences to compered with control which showed only 12%. was quite low, while there were no significant differences between the treatments.



Treatments / Time (min)

Fig. 6: The effects of different duration of sulfuric acid (98 %) on germination percentage (GP) of *Alhagi graecorum* seeds, Values are means \pm (n = 4) (P < 0.05)

leguminous family have hard and impermeable seed coats caused by physically dormancy ^[25]. Generally, legume seeds exhibit hard seededness resulting in dormancy. Several studies have been conducted on legume germination using different seed coat presowing treatments. The function of the seed coat is to protect the embryo and endosperm from desiccation, mechanical injury, unfavorable temperatures and attacks by bacteria, fungi and insects ^[27]. Seeds of the fabaceae family exhibit dormancy because of hard testa impermeable to water and gases ^{[15],[17],[28]}. The results of this study are in agreement with those reported by ^[29] and ^[17] showing that,

many species of the fabaceae family such as Lupinus spp. Seeds exhibit dormancy that is primarily due to water impermeability of the seed's coat. Scarification of Texas bluebonnet (Lupinus texensis Hook) seeds with sulfuric acid for 30 to 60 min improved seedling emergence.^[27] reported that, mechanical scarification of intact seeds significantly (P < 0.05) increased germination percentage and recorded the highest germination percentage among all treatments during the entire germination period followed by immersion of intact seeds in H₂SO₄ for 30 min treatments. The concentrated sulfuric acid treatment has been widely used to improve seed germination of several hard seed coat species [30]. Acids (HCl, HNO3, and H₂SO₄) have been widely used for breaking dormancy of many hard seed coat species, such as European milkvetch (Astragalus hamosus L.), blackdisk medick (Medicago orbicularis (L.) Bartal. [31] and Albizia spp. [30], [17] indicated that, the best treatment to remove hard seed dormancy causing the highest germination percentage was seed scarification with H₂SO₄ and sandpaper. It was observed that for archer and perennial soybeans, using H2SO4 immersion was the method that had the highest percentage of germination, so the best method to effect the removal of dormancy [32]. Previous work on Parkia biglobosa [33], Enterolobium contortisiliquum [34], Rhynchosia capitata [17] also showed that soaking of seeds of these plants in H₂SO₄ can break dormancy and increase germination percentage. According to ^[35], the perennial soybean germination without scarification has a ranging between 7% and 24%, with immersion in H₂SO₄ there was a 100% increase in the germination of their seeds compared to the control which showed only 26% germination. In seeds Piptadeniamoniliformis found higher percentages of germination in treatments subjected to immersion in sulfuric acid for 20, 25 and 30 min [36]. According to [37], scarification of seed coat with acids such as H₂SO₄ usually leads to the elimination of exogenous dormancy. These findings are consistent with our findings.

Mean daily germination (MDG), is an index of the daily germination rate. In (Fig.7) the results exhibited that, there were highly significant differences in mean daily germination of germination percentage on the third day of sowing in the treated seeds, 10, 20, 30 and 40 mins compared with control. It was the fastest and most effective in germination achieved 84, 89, 90, 92 % respectively, while in the control was 0 %.

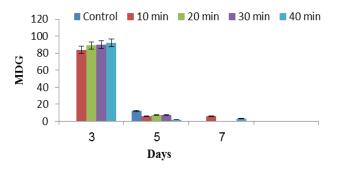


Fig. 7: The effects of different duration of acid (98 %) on mean daily germination (MDG) values of *Alhagi graecorum* seeds (P < 0.05)

Mean germination time (MGT) is an accurate measure of the time taken for a lot to germinate, but does not correlate this well with the time spread or uniformity of germination. It focuses instead on the day when most germination events occurred ^[26], it is interpreting the time taken to achieve the most germination of seeds.

The results in (Fig. 8) revealed that, the germination time (MGT) was decreased in all treatments of H_2SO_4 , 10 min, 20 min, 30 min

and 40 min were statistically the same but significantly different from the control

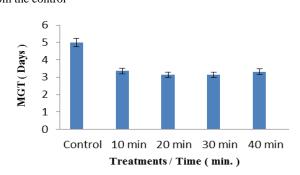
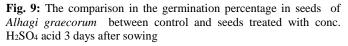


Fig. 8: The effects of different duration of sulfuric acid (98 %) on mean germination time (MGT) values of *Alhagi graecorum* seeds (p < 0.05).

The minimum MGT recorded in treated seeds in 20 min and 30 min were 3.14 and 3.14 days respectively. These results are consistent with ^[38], reported that the lowest MGT values in *Medicago scutellata* and *Medicago polymorpha* species were observed in the seeds treated with H₂SO₄, treatments for 20, 30 min revealed that, H₂SO₄ were adequate to break the hard seed coat of *Alhagi graecorum* seeds in order to induce germination. The results of this experiment confirmed that the *Alhagi graecorum* seeds were in a dormant state. Scarification with H₂SO₄ overcame seed dormancy and increased the germination percentage (Fig.9). It can be stated that the most effective chemical scarification method in breaking seed dormancy of *Alhagi graecorum* is H₂SO₄ treatments. GP, MDG and MGT values at the end of all treatments were found to be quite high, while MGT value was quite low when compared to the control (Fig. 10).





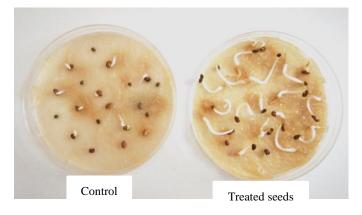


Fig. 10: The comparison in the germination percentage in seeds of *Alhagi graecorum* between control and seeds treated with conc. H₂SO₄ acid 5 days after sowing

Conclusion

The results of the current study demonstrated that seeds of *Alhagi-graecorum* exhibits dormancy imposed by a hard seed coat. Softening of the seed coat by scarifying with concentrated H₂SO₄ significantly increased seed germination percentage (GP), mean daily germination (MDG) and reduced mean germination time (MGT). scarification with H₂SO₄ was the fastest and the most effective dormancy breaking methods for *A. graecorum* seeds.

Therefore, I recommend to use concentrated sulfuric acid when germinating the seeds of the native species of *Alhagi graecorum*.

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References

- [1]- Galíndez, G., Ceccato, D., Malagrina, G., Pidal, B., Chilo, G., Bach, H., Fortunato, R. and Ortega-baes, P. (2016). Physical seed dormancy in native legume species of Argentina. Bol. Soc. Argent. Bot. 51 (1): 73-78.
- [2]- Singh,V. P, Yadav, B. and Pandey,V. B. (1999). Flavanone glycosides from *Alhagi pseudalhagi*. Phytochemistry, **51**: 587-590.
- [3]- Boulos L (2000). Flora of Egypt, Al Hadara Publishing, Cairo, Egypt. pp 287.
- [4]- Al-Massarani1, S. and El Dib, R. . (2015). *In vitro* evaluation of cytotoxic and antimicrobial potentials of the Saudi traditional plant *Alhagi graecorum* boiss. *Pak. J. Pharm. Sci.*, Vol. 28, No.3,1079 -1086.
- [5]- Emmanuel, A. and Olayinka, O. (2018). Scarification of Exotic and Indigenous Plant Seeds in Nigeria: Effect on Dormancy and Germination. http://dx.doi.org/10.1101/354993
- [6]- Marchetti, R. (2012). Evaluation of Four Treatments to Break Seed Dormancy of *Sunflower Inbreds*. Master of Science in Agriculture and Natural Resources Degree. University of Tennessee at Martin.
- [7]- Baskin C. C. and Baskin, J. M. (1998). Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination. San Diego: Academic Press.
- [8]- Hilhorst, H. W. M. (2007). Definitions and hypotheses of seed dormancy. In: Bradford K. & Nonogaki H. (eds.), Seed Development, Dormancy and Germination, Annual Plant Reviews, Vol. 27. Sheffield: Blackwell Publishing. pp.50– 71.
- [9]- Leo, J. (2013). The Effect of Cold Stratification on Germination in 28 Cultural Relict Plant Species With the Purpose of Establishing Germination Protocols. Swedish University of Agricultural Sciences.
- [10]-Bewley J. D. (1997). Seed Germination and Dormancy, The Plant Cell, vol. 9, pp. 1055–1066.
- [11]-Baskin C. C. and Baskin J.M. (2005). Seed dormancy in wild flowers. In: McDonald M.B. and Kwong F. Y. (eds.), Flower Seeds: Biology and Technology. Wallingford: CABI Publishing, pp. 163–185.
- [12]-Kucera, B., Marc Alan, C. and Gerhard, L.M. (2005). Plant hormone interactions during seed dormancy release and germination. Seed Science Research, 15, 281–307.
- [13]-Teimouri, M.S, koocheki, A. and Mahallati, M.N. (2013). Seed germination and breaking of seed dormancy techniques for endemic Hymenocrater platystegius Rech.f. of Khorasan Razavi province, Iran. Intl J Agri Crop Sci. Vol., 6 (12), 885-889.
- [14]-Finch-Savage, W.E. and Leubner-Metzger, G. (2006). Seed dormancy and the control of Germination. New Phytologist, 171: 501–523.
- [15]-Pallavi, H. M., Vishwanath, K., Harish, B. S., Prashanth, Y. and Manjunath, T.(2014). Seed treatments to break seed dormancy and Standardization of viability test procedure in *Abrus precatorious*. Journal of Medicinal Plant Research. Vol. 8 (4), 229-236.

- [16]-Graeber, K., Kazumi, N., Emma, M., Gerhard, LM. and Wim, S. (2012). Molecular mechanisms of seed dormancy. Blackwell Publishing Ltd, Plant, Cell and Environment.
- [17]-Ali, H.H., Asif, T., Tanveer, A., Nadeem, M.A. and Asghar, H.N. (2011). Methods to break seed dormancy of *Rhynchosia capitata*, A summer annual weed. Chilean journal of agricultural research 71(3): 483 – 48.
- [18]-Gama-Arachchige, N.S., Baskin, J.M., Geneve, R.L. and Baskin, C.C. (2010). Identification and characterization of the water gap in physically dormant seeds of Geraniaceae, with special reference to Geranium carolinianum. Annals of Botany 105: 977–990.
- [19]-Yildiztugay, E. and Kucukoduk, M. (2012). Dormancy breaking and germination requirements for seeds of *Sphaerophysa kotschyana* Boiss. Journal of Global Biosciences. Vol. 1, pp. 20-27.
- [20]-De Paula, A.S., Carolina, M.D., Maria, T. S. P. and Marisa, S. (2012). Breaking physical dormancy of *Cassia leptophylla* and *Senna macranthera* (Fabaceae: Caesalpinioideae) seeds: water absorption and alternating temperatures. Seed Science Research. Cambridge University Press. 22, 259–267.
- [21]-De Souza, T.V., Voltolini, C.H., Marisa Santos, M. and Paulilo, M. T.S. (2012).Water absorption and dormancybreaking requirements of physically dormant seeds of *Schizolobium parahyba* (Fabaceae – Caesalpinioideae). Seed Science Research, Cambridge University Press, pp. 1-8.
- [22]-Baskin J.M. and Baskin C.C. (2004). A classification system for seed dormancy. Seed Sci. Res. 14 (1): 1–16.
- [23]-Arowosegbe, S. (2016). Studies on methods of breaking seed dormancy and germination enhancement in *Senna alata* (L.) Roxb., A Plant with great medicinal value. International Research Journal of Natural Sciences. Vol.4, No.2, pp.31-40.
- [24]-Scott S.J., Jones, R.A. and Williams, W.A. (1984). Review of data analysis methods for seed germination. Crop Sci. 24: 1192-1199.
- [25]-Mirzaeil, M., Moghadam, A. R.M., Ardebili, Z.O. (2013). The induction of seed germination using sulfuric acid, gibberellic acid and hot water in *Robinia pseudoacacia* L. Intl. Res. J. Appl. Basic. Sci. Vol., 4 (1), 96-98.
- [26]-Kader, M. A. (2005). A Comparison of Seed Germination Calculation Formulae and the Associated Interpretation of Resulting Data. Journal & Proceedings of the Royal Society of New South Wales, Vol. 138, 65–75.
- [27]-Naim, A.H., El Hadi, A.H. and Ahmed, F..E. (2015). Evaluation of different pre-sowing seed treatments to break dormancy of *Crotalaria senegalensis*, a famous rangeland forage in Sudan. Asian J. Plant Sci. Res., 5(10):16-21.
- [28]-Shaik, S., Dewir, Y. H, Singh, N. and Nicholas, A. (2008). Influences of pre-sowing seed treatments on germination of the cancer bush (*Sutherlandia frutescens*), a reputed medicinal plant in arid environments. Seed Sci. Technol. 36(3):795-801.
- [29]-Davis, T. D., George, S. W., Upadhaya, A. and Parsons, M. .(1991). Improvement of seedling emergence of *Lupinus texensis* following seed scarification treatments. Journal of Environmental Horticulture 9:17-21.
- [30]-Tigabu, M., and Oden, P. C.. (2001). Effect of seed scarification, gibberellic acid and temperature on seed germination of two multipurpose *Albizia* species from Ethiopia. Seed Science and Technology 29:11-20.
- [31]-Patane, C., and F. Gresta. (2006). Germination of Astragalus hamosus and Medicago orbicularis as affected by seed coat dormancy breaking techniques. Journal of Arid Environment 67:165-173.
- [32]-De Morais, L.F., Almeida, J.C.C., Deminicis, B.B., de Pádua, F.T., Morenz, M.J.F., de Abreu, J.B.R., Araujo, R.P. and de Nepomuceno, D.D. (2014). Methods for Breaking Dormancy of Seeds of Tropical Forage Legumes. American Journal of Plant Sciences, 5, 1831-1835. http://dx.doi.org/10.4236/ajps.2014.513196.
- [33]-Aliero, B.I. (2004). Effects of sulfuric acid, mechanical scarification and wet heat treatments on germination of seeds

of African locust bean tree. *Parkia biglobosa*. African journal of Biotechnology 3: 179-181.

- [34]-Malavasi, U.C. and Malavasi, M. (2004). Dormancy breaking and germination of *Enterolobium contortisiliquum* (Vell.) Morong seed. Brazilian Archives of Biology and Technology. 47:851-854.
- [35]-Neme, A.N. (1963). Perennial Soybean Seeds. Results of Scarification and Duration of Power Twinning. Bragantia: Scientific Bulletin of the Agricultural Institute of the Sao Paulo State, 22, 785-791.
- [36]-Azeredo, G.A., Paula, R.C., Valeri, S.V. and Moro, F.V. (2010) Overcoming Seed Dormancy in *Piptadenia moniliformis* Benth. Brazilian Journal of Seeds, 32, 49-58.
- [37]-Nadjafi, F., Bannayana, M., Tabrizia, L. and Rastgoo, M. (2006). Seed germination and dormancy breaking techniques for *Ferula- gummosa* and *Teucrium polium*. Journal of Arid Environment. 64: 542-547
- [38]-Khaef, N., Sadeghi, H. and Taghvaei, M. (2011). Effects of new strategies for breaking dormancy of two annual medics (*Medicago scutellata* and *Medicago polymorpha*), American-Eurasian J. Agric. & Environ. Sci., 11(5), 626-632.